Tardigrades of Fish Creek Provincial Park, Alberta, Canada: A Preliminary Survey

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Tardigrades are cosmopolitan animals with a patchy distribution record, largely due to the limited collection history for this phylum rather than actual rarity. Most Canadian records are from near coastal areas, with a few notable exceptions, most recently one study of tardigrades in central Alberta. The present study reports the occurrence of 12 species or species groups in Fish Creek Provincial Park in southern Alberta. Eight of these taxa are new records for Alberta, and two (*Diphascon (Diphascon) granifer* and *Echiniscus granulatus*) are new records for Canada.

Key Words: Tardigrada, survey, meiofauna, Alberta.

Tardigrades (phylum Tardigrada), also commonly known as water bears, are microscopic invertebrates (normally < 800 µm) (Kinchin 1994) found in ecosystems from the Arctic and the Antarctic to the tropics, on sandy beaches, the ocean bottom, in shallow ponds, and, most famously, in mosses (Kinchin 1994; McInnes 1994). While all tardigrades in their active state are dependent on a film of water and so may be considered aquatic, for convenience those from non-submerged habitats may be described as terrestrial (Ramazzotti and Maucci 1983).

Their most widely recognized characteristic is remarkable resilience, including an ability to survive long periods without water by entering an anhydrobiotic stage in which they crumple down to resemble a speck of dust, surviving for up to 10 years in this desiccated state before being rehydrated and resuming normal life (Jönsson and Bertolani 2001; Bertolani et al. 2004). They have been shown to resist extremes of heat (Doyère 1842; Rahm 1921) and cold (Becquerel 1950; Hengherr et al. 2009), radiation (May et al. 1964), high pressure, and vacuum (Seki and Toyoshima 1998)—indeed, they are the only animals so far to have been deliberately exposed to the vacuum and radiation of space, which they survived (Jönsson et al. 2008).

Fish Creek Provincial Park (50°55'N, 114°3'W) is a natural area in Calgary, Alberta, at an elevation of approximately 1048 m (Figure 1). The area has long, dry, cold but highly variable winters and short, moderately warm summers; the yearly mean temperature is 4.1°C and the mean annual precipitation is 412.6 mm (National Climate Data and Information Archive 2010*). At 1348 ha, Fish Creek Provincial Park is one of Canada's largest urban parks, and it is almost entirely enclosed within the city (Friends of Fish Creek 2010*).

Fish Creek Provincial Park is at the northwestern edge of the Prairie ecozone, and it contains grasslands, Trembling Aspen (*Populus tremuloides*) woods, riparian forest, White Spruce (*Picea glauca*) forest, and natural and engineered wetlands (Robertson 1991; Friends of Fish Creek 2010*). Trembling Aspens tend to grow at the edge of floodplain terraces, along the tops of north-facing slopes, and in ravines, while riparian vegetation is dominated by Balsam Poplars (*Populus balsamifera*) and willows (*Salix* spp.) (Pinel 1980). White Spruce are located on north-facing slopes and floodplains between the Trembling Aspen and riparian vegetation in the western end of the park (Pinel 1980). Grasslands occur on the floodplain and drier southfacing slopes (Pinel 1980).

Only one previous survey of terrestrial tardigrades has been carried out in Alberta (Boeckner and Proctor 2005). Fish Creek Provincial Park is an ideal location in which to begin a more widespread survey. It not only includes a good mixture of ecosystem types, but it also contains both natural and disturbed areas appropriate for longitudinal studies of anthropogenic impacts on tardigrades. Relatively few studies of the colonization of newly altered environments by tardigrades have been performed (Uhía and Briones 2002; Hohberg 2006), so the colonization of the engineered wetlands recently completed in the Park to treat stormwater runoff from the surrounding communities will be of particular interest (Friends of Fish Creek 2009*). In their cryptobiotic state, tardigrades are easily dispersed by passive transport (Nelson et al. 2009); therefore, terrestrial tardigrades should be fairly early colonizers of disturbed areas, particularly areas with erratic periods of moisture and temperature. Truly aquatic tardigrades, which lack a desiccated "tun" stage with its presumed wind-dispersal capacity, are likely to be less easily dispersed (Nelson et al. 2009).

There is a general need for wider surveys of the distribution of this phylum; the majority of the more than 1000 species described have been found in only one region, or even in only a single sampling site (McInnes

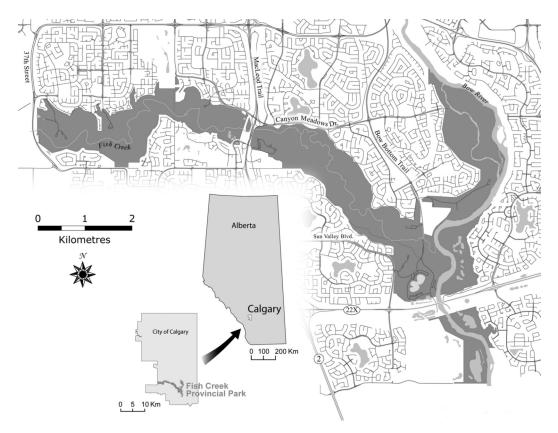


FIGURE 1. Fish Creek Provincial Park, Calgary, Alberta, Canada (Alberta Tourism, Parks and Recreation, Canmore)

1994; Pilato and Binda 2001). This is likely due as much to the patchiness of tardigrade surveys as to any patchiness in tardigrade distribution.

Previous studies that concentrated on or included Canadian tardigrades are usefully summarized in Bateman and Collins (2001), updated by Boeckner et al. (2006). Since the preparation of these two lists, two additional studies have been published (Boeckner and Proctor 2005; Collins 2010). The majority of work has been performed on so-called terrestrial tardigrades, which inhabit mostly moss, lichen, and leaf litter, with only one study, of mountain lakes, including tardigrades of permanent water bodies (Anderson and de Henau 1980).

Methods

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In total, 73 samples were collected, and GPS coordinates and location descriptions were recorded. The sampling and processing procedure of Nelson et al. (2009) was used. Samples of dry moss (~250 mL), lichen (~1 mL), and leaf litter (~250 mL) were collected by hand and stored up to one year, but normally less than two weeks, in paper bags in a sealed specimen cabinet. This procedure will have selected against species with little capacity for anhydrobiotic survival. Samples of aquatic sediment (~125 mL) were collected in jars and processed within six hours.

Dry samples were soaked in bottled water (Culligan) for 16–24 hours, agitated, and filtered through stacked sieves (1.4 mm, 75 µm, and in some cases 45 µm). The 75 µm and 45 µm sieves were backwashed, with the collected material immediately fixed in boiling 95% reagent alcohol. In cases of heavy burden with inorganic matter, a modification of the procedure of Hallas (1975) was followed: samples were allowed to settle for 5 minutes, the precipitate was re-suspended in 50% sucrose and centrifuged for 5 minutes at 1200 × g, and then the supernatant was re-filtered through the finest sieve and collected by backwashing with 80% alcohol. Aquatic samples were briefly stirred and allowed to settle to remove the most dense sediment, and then the liquid was decanted and filtered as above.

Samples were examined with a stereomicroscope at 30×, and specimens were mounted in polyvinyl alcohol (Bioquip 6371A) for observation with a Leica DME compound microscope using brightfield and phase contrast. Specimens were observed immediately in order to record perishable characters such as colour and eyespots, then the specimens allowed to decolourize for two weeks and were re-observed for internal

	Substrate		
	Moss	Lichen	Leaf litter
Number of samples that contained tardigrades	14	9	7
(total number of samples in parentheses)	(29)	(12)	(14)
Diphascon (Adropion) cf. prorsirostre		2	
Diphascon (Diphascon) granifer			3
Diphascon (D) pingue gr.	4		
Diphascon (D) sp.		1	
Echiniscus granulatus	1		
Hypsibius convergens			4
Isohypsibius lunulatus	25		
Isohypsibius cf. prosostomus	40		
Isohypsibius tuberculatus gr.			2
Macrobiotus cf. harmsworthi	11	1	
Milnesium tardigradum gr.	10	16	4
Paramacrobiotus cf. richtersi	8		5
Ramazzottius oberhaeuseri gr.	47	47	3
Unidentified to this level	141	7	22

TABLE 1. Number of tardigrades, by species in each substrate, recovered from Fish Creek Provincial Park. Abbreviations used: cf. = provisional identification to species; gr. = group of closely related species; sp. = unidentified species of subgenus.

morphology. Permanent mounts of all specimens are stored at St. Mary's University College and are available for review.

Identification of specimens was based primarily on the keys of Ramazzotti and Maucci (1983); Dastych (1988); Nelson et al. (2009); Pilato and Binda (2010). Taxonomy and nomenclature were confirmed with the most recent checklists (Guidetti and Bertolani 2005; Degma and Guidetti 2007; Degma et al. 2010*).

Results

Tardigrades were extracted from fewer than half (33 of 73) of the samples examined. Of 409 tardigrades extracted, 234 could be identified to species or speciesgroup level. As many tardigrades can be identified to species only by differences in egg morphology and few eggs were recovered from these samples, many of the specimens are identified only to subgenus or to a group of closely related species (Table 1). There were a minimum of 1 to a maximum of 5 species (mean 1.7) per positive sample. No tardigrades were found in 11 aquatic samples from engineered ponds, while 3 of 7 samples from Fish Creek or a natural pond yielded only a total of 5 specimens in such poor condition that they could not be readily identified (one *Isohypsibius* sp., two Hypsibiidae, and two other Eutardigrada).

Two species, *Diphascon* (subgenus *Diphascon*) granifer (Figure 2) and *Echiniscus granulatus* (Figure 3), are new records for Canada (Bateman and Collins 2001; Boeckner et al. 2006). *Echiniscus granulatus* is a widespread species found in several geographic regions (McInnes 1994), and both species have previously been recovered in North America (McInnes 1994; Bartels and Nelson 2007). *Diphascon* is a new genus for Alberta, represented here by both subgenera *Diphascon* (*D. granifer* and *D. pingue* gr.) and *Adropi*on (*D. cf. prorsirostre*). *Isohypsibius lunulatus*, *I. cf.*

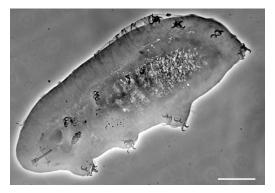


FIGURE 2. *Diphascon (Diphascon) granifer* beginning to shed (note duplicated claws and cuticle pulling away). Phase contrast. Scale bar = 50 μm.

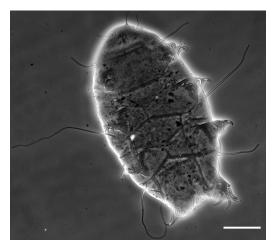


FIGURE 3. *Echiniscus granulatus*. Phase contrast. Scale bar $= 50 \mu m$.

prosostomus, and Macrobiotus cf. harmsworthi have not previously been recorded in Alberta.

Nine of 12 lichen samples contained tardigrades, principally of the *Ramazzottius oberhaeuseri* group (47 individuals) and the *Milnesium tardigradum* group (16). Tardigrades were recovered from half of moss and litter samples (14 of 29 and 7 of 14, respectively). In moss, the *Ramazzottius oberhaeuseri* group (47) was most abundant, followed by *Isohypsibius* cf. *prosostomus* (40) and *Isohypsibius lunulatus* (25).

Discussion

In the absence of eggs, it was not possible to identify the collected Ramazzottius oberhaeuseri gr. specimens to species, but close observation suggests the presence of at least three distinct species, based on cuticular coloration and sculpture. Differences in morphology in the macroplacoids and cuticle also suggest that the specimens identified as Macrobiotus cf. harmsworthi and Paramacrobiotus cf. richtersi (listed in Boeckner et al. (2006) by its former nomenclature, Macrobiotus richtersi) may include multiple closely related species. Further collection and morphometric analysis should permit a more precise determination. Milnesium, long considered a monospecific genus, now has 14 new species described in the last two decades (Degma et al. 2010*). It is represented here by the species-group Milnesium tardigradum gr.

Of the examined substrates, lichen was most likely to harbour tardigrades, primarily Ramazzottius oberhaeuseri gr. and Milnesium tardigradum gr., but leaf litter had the most diverse assemblages, with a mean of 2.1 species per positive sample compared to 1.6 for lichen. The two most diverse samples, with 5 and 4 species, were both from leaf litter. This may reflect the diverse nature of leaf litter itself, which can be considered a vertical set of different microhabitats with shallow soil at the bottom. It would be worthwhile in future studies to differentiate leaf litter by the dominant type of vegetation. Trembling Aspen and White Spruce, for example, are expected to produce litter differing in numerous ways. On the other hand, moss yielded the greatest numbers of tardigrades: 20.5 per positive sample versus 8.2 for lichen and 6.1 for leaf litter.

Sample sizes were too small to make strong statements as to species distribution at present, particularly as samples from different substrates were not standardized for mass, but these figures agree generally with previous work, which has shown greater abundance and species diversity in moss and leaf litter than in lichen (Miller et al. 1996; Avdonina 2006; Bartels and Nelson 2007).

Overall, the number of tardigrades recovered was not particularly high, and many samples contained no tardigrades at all. Certainly, yields were considerably lower than those of Boeckner and Proctor (2005), who sampled moss on Black Cat Mountain in central Alberta. It is possible that the more urban nature of Fish Creek Provincial Park contributes to this discrepancy; it is recognized that tardigrade distribution is sensitive to, and a possible biomarker for, air quality (Moly de Peluffo et al. 2006). Fish Creek Provincial Park is a long, narrow park confined to the valley around Fish Creek. Since the Park is largely surrounded by subdivisions and is crossed by two major highways, air quality may be a factor limiting tardigrade distribution.

As expected, no tardigrades were recovered from the water of the recently created wetlands; repeat sampling in future years will be necessary to track colonization of these new environments as well as to make comparisons with similar natural wetlands in the region. It will be interesting to compare these results with observations from a wider area in southern Alberta. Based on reports of seasonality in freshwater and terrestrial tardigrades (several summarized in Kinchin 1994), it would also be useful to look for seasonal variation in tardigrade distribution while monitoring changes in the population of Fish Creek Provincial Park.

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